

How Desert Succulents Keep Their Stomata Closed by Day without Starving for Air

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Abstract

Desert succulents face one of the sharpest physiological dilemmas in the plant kingdom they live in places where opening their stomata in the day would cause near lethal water loss, yet they still need carbon dioxide to photosynthesize and just keep going. The workaround they evolved is Crassulacean Acid Metabolism, or CAM photosynthesis, a biochemical trick so neat it has popped up on its own in more than 60 plant families. Instead of pulling in CO₂ during daylight, like most plants do, CAM plants open their stomata at night, grab carbon dioxide and convert it into organic acids, tuck those acids away inside vacuoles, and then, during daytime, release the CO₂ and refix it while the stomata stay tightly closed. Here we look at the whole mechanistic sequence of CAM photosynthesis, how succulents handle the constant bargain between carbon gain and water conservation, and the ecological plus evolutionary backdrop that likely shaped this adaptation. The discussion also covers physiological flexibility across CAM plants, why vacuolar storage capacity matters so much, and what CAM photosynthesis might imply for food and water security research as climate change speeds up. When you understand how these plants sidestep daytime starvation without opening their stomata, you start to see some of the most sophisticated terrestrial plant biochemistry.

Keywords: CAM photosynthesis, stomatal regulation, malate storage, crassulacean acid metabolism, desert succulents, water use efficiency

I. Introduction

Imagine being made to hold your breath all day but still, somehow, figuring out how to eat. That's kinda the weird deal desert succulents face every time the sun comes up. Most plants? They do photosynthesis and gas exchange when it's bright outside—stomata open in the morning, CO₂ comes in, sunlight powers the Calvin cycle, sugars get built. Pretty straightforward. However, in a place where midday temperatures can jump past 50°C and the relative humidity can fall under 10%, opening stomata during the day is basically like volunteering to dry out. The water loss through those little pores isn't some gentle leak either it's more like catastrophic, quickly and without mercy.

So, succulents went with a different rhythm. They don't really open their stomata during daytime, not when it counts. Instead, they handle gas exchange at night, when it's cooler and the difference in vapor pressure between the leaf interior and the dry desert air becomes much less brutal. They take in CO₂ in the dark, then lock it inside their cells as malic acid, and later they release that stored carbon during daylight to run their photosynthetic machinery—still keeping stomata shut.

This approach is kinda known as Crassulacean Acid Metabolism, or CAM, named after the Crassulaceae family (the stonecrops and jade plants) where it was first shown in biochemical terms. But it's not only, like, a "that family thing" because CAM has been reported across more than 60 plant families, in roughly 7% of studied vascular plant species, including cacti, agaves, bromeliads, orchids, and even some aquatic plants (Silvera et al., 2010). The fact that it evolved so many separate times, on its own, tells you something pretty direct—when the alternative is just dying under heat, evolution seems to find a workaround.

This article walks through exactly how CAM works, why it handles the starvation versus dehydration dilemma so effectively, what its metabolic costs, plus constraints really are, and why researchers are getting more and more interested in it as a kind of blueprint for engineering more drought-tolerant crops in a warming world.

II. The Problem That CAM Solves

2.1 Water Loss Through Open Stomata

To appreciate CAM, you sort of need to get why stomata are such a trouble, in hot dry environments. Stomata are microscopic openings on leaf surfaces, and they're flanked by guard cells, that control the opening and closing. When the stomata open so CO₂ can come in, well they inevitably let water vapor out. The whole push for that loss comes from a difference in water vapor concentration, between the leaf interior which is humid, and the surrounding air which is drier — plant physiologists call it vapor pressure deficit (VPD).

In desert conditions, VPD is extreme. A succulent leaf in the Sonoran Desert on a July afternoon might have an internal relative humidity close to 100% , while outside air sits at 15% humidity or even less. That gap keeps pulling water out of the leaf at rates that no stash of stored water can keep up with for long. Research by Nobel (1988) showed that the water use efficiency of CAM plants — basically grams of carbon fixed per kilogram of water lost — can be about three to five times higher than that of C_3 plants under comparable conditions. Most of that benefit comes down to when stomata are open, not really whether they open.

By closing stomata during the day, the plant avoids the most punishing stretch of water demand. Desert heat often tops out early afternoon, VPD rises with it, and wind (that strips away the humid boundary layer along leaf surfaces) frequently does the same. A CAM plant that keeps stomata shut through all of that can make it through situations that would normally desiccate a standard C_3 plant, quite fatally.

2.2 The CO_2 Supply Problem

So, here's the catch, photosynthesis really runs on CO_2 . The Calvin cycle—where carbon is actually fixed into sugars—can't go on without a steady feed of that gas. If you keep your stomata shut all day, then you basically stop the supply, and photosynthesis grinds to a halt. It's kind of a dilemma, you know. Plants need light and CO_2 at the same time so the Calvin cycle can work. But the light shows up in the daytime, and the safest, most reliable time to grab CO_2 is usually at night.

CAM breaks that constraint by separating, or decoupling, gas exchange from carbon fixation across the day-night cycle. In plain terms, the plant gathers CO_2 at night and stores it. Then when daytime comes, it pulls from those stores to power the Calvin cycle. The gas exchange and the carbon-fixing steps happen at different times, but they're still biochemically connected, kind of like filling a water tank at night and then using that water during the day.

III. The Biochemistry of CAM: A Four-Phase System

Researchers have kind of divided the CAM cycle into four phases, each one supposed to show a different slice of metabolic activity across that 24-hour stretch. This framework, worked out mostly through Osmond (1978) and then later refined, has ended up sticking around as the usual way people describe how the system basically runs.

Phase I: Nighttime CO_2 Fixation

At night the guard cells kind of respond to darkness and those dropping temperatures by opening up the stomata, basically. CO_2 then diffuses into the mesophyll cells, of this succulent leaf. Once inside, an enzyme called phosphoenolpyruvate carboxylase (PEPC) catches the CO_2 and combines it with phosphoenolpyruvate (PEP) so it forms oxaloacetate, a sort of four-carbon compound. Oxaloacetate is then quickly reduced into malate and that malate gets pumped into big central vacuoles where it piles up as malic acid all through the night.

The acidification of vacuoles is not just a metaphor, it is literal and you can measure it. CAM plants show this clear diel, 24-hour swing in leaf acidity— they can taste sour in the morning, and much less so by evening. People who watched plants closely noticed that pattern long before anyone had the biochemistry worked out in detail. The ability of a plant's vacuoles to store malic acid puts a ceiling on the amount of CO_2 it can fix at night, and that ceiling later limits the plant's overall daily carbon gain.

Phase II: The Dawn Transition

As dawn breaks, and light starts to filter into the leaf, both carboxylation routes are kind of active at the same time, just briefly. PEPC keeps on fixing atmospheric CO_2 while, at the same time decarboxylation of malate starts, and that begins to provide CO_2 for the Calvin cycle. Stomata might stay partly open during this quick handoff, taking in a bit of direct atmospheric CO_2 before the daytime heat really ramps up. Phase II is short, usually one to two hours, but it is a sort of overlap point between the nocturnal and diurnal metabolic programs.

Phase III: Daytime Decarboxylation and Carbon Refixation

This is where the real magic happens, stomata close a bit too tightly when temperatures rise and the VPD goes up. Inside the leaf, malate starts moving out of the vacuoles and into the cytoplasm, where it gets decarboxylated, so it is stripped of its saved CO_2 . The exact enzyme doing that depends on the species, yes, not everyone uses the same trick. Some plants rely on NAD-malic enzyme, others use NADP-malic enzyme, and still others swap to phosphoenolpyruvate carboxykinase. In every case they release CO_2 into the cell interior. That CO_2 then builds up to concentrations way above atmospheric levels, but only inside the leaf. After that, the CO_2 gets captured by RuBisCO and pushed into the Calvin cycle, which runs on light energy from the photosystems sitting in the chloroplasts. The critical part is that it all plays out while stomata stay shut, the CO_2

is recycled within rather than pulled in from outside. Basically, the plant is doing photosynthesis using stored reserves, burning through its biochemical savings from the night before, kind of a quiet metabolic rerun. As shown in Figure 1, the diel fluctuation in leaf malate concentration mirrors this four-phase cycle almost exactly — rising steeply through the night, peaking at dawn, declining sharply through the morning and midday as decarboxylation proceeds, and hitting a daily minimum by late afternoon when malate stores are largely exhausted.

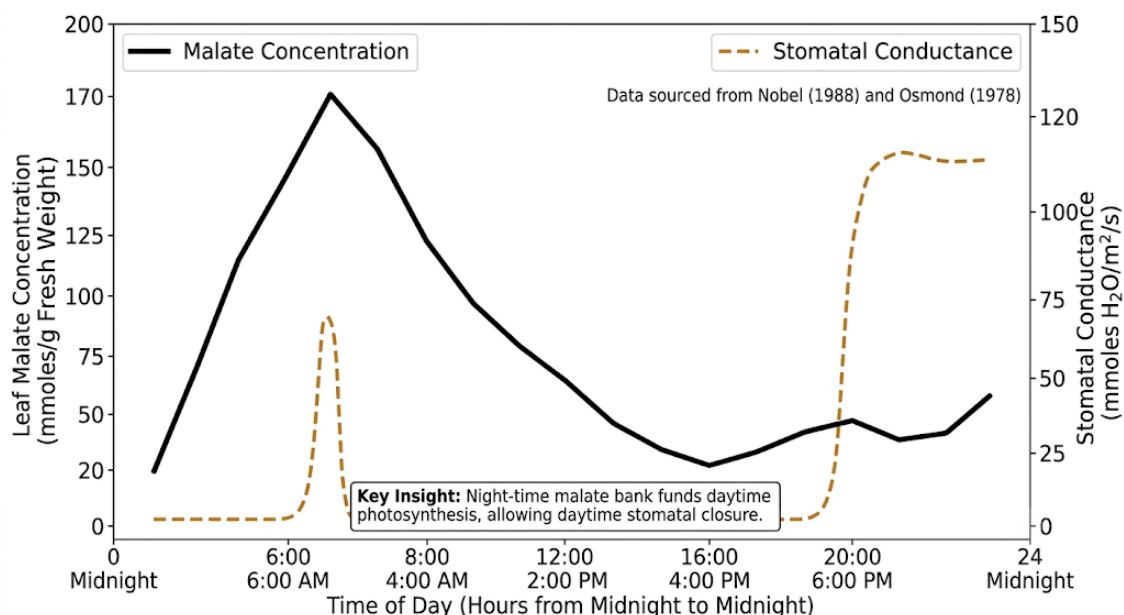


Figure 1: Diel Fluctuation in Leaf Malate Concentration and Stomatal Conductance in a CAM Succulent Over 24 Hours

This figure shows a dual axis line graph, you know tracking leaf malate concentration (millimoles per gram fresh weight, left y-axis) and stomatal conductance (millimoles of water vapor per square meter per second, right y-axis) over a full 24-hour span for a “representative” CAM succulent, (*Agave deserti*) under natural desert settings. The x-axis, just time, runs from midnight straight to midnight again.

The malate concentration line goes up pretty quickly starting at midnight, keeps climbing until about 6:00 AM, then it hits a peak around dawn. After that it keeps tapering off through the morning and into midday, and it bottoms out near 16:00. On the other hand, the stomatal conductance line behaves kinda like the opposite pattern—it starts climbing at dusk, stays high through the night, then drops sharply at dawn. During the daylight hours it stays close to zero, which basically highlights that when stomata shut during daytime the internal CO₂ supply still gets covered, likely via malate breakdown. In other words, you see inverse coupling between daytime stomatal closure and CO₂ availability from that internal pool.

The data are credited to Nobel (1988), *Environmental Biology of Agaves and Cacti*, Cambridge University Press, along with Osmond (1978) from *Annual Review of Plant Physiology*. The main take away is pretty straightforward: the malate bank at night essentially underwrites the daytime photosynthetic effort, so daytime stomatal opening becomes unnecessary, or at least it’s dramatically reduced.

Phase IV: Late Afternoon CO₂ Uptake

By late afternoon, the malate reserves are mostly depleted. Temperatures start dropping, VPD eases, and a few CAM species begin to reopen their stomata, a bit cautiously, to snatch a small amount of atmospheric CO₂ straight from the air. That late day window gives a little extra carbon pickup before nightfall kicks in and the full Phase I routine has to restart. Not every CAM plant shows a clear Phase IV though, it depends on species, season, and water condition, but in well-watered individuals it acts like a modest add-on to nocturnal fixation.

IV. How Succulents Are Built to Support This Biochemistry

4.1 Vacuolar Volume as a Limiting Factor

The whole CAM strategy kind of hangs on having enough vacuolar storage capacity to stash meaningful amounts of malic acid overnight. So the succulence of CAM plants isn’t just a thing about keeping water reserves, it’s more like functionally tied to the CAM system itself. Thick, water-filled leaves and stems

have big mesophyll cells with proportionally large vacuoles. And those vacuoles act as the storage tanks for nocturnal CO₂ fixation, pretty directly.

Nobel (1988) showed a strong positive correlation between tissue succulence and CO₂ uptake capacity across multiple CAM species. That basically confirms that the more succulent the plant is, the more carbon it can fix per night. This connection helps explain why CAM plants often lean toward fleshy architectures, because the succulence isn't accidental it's structurally connected to the metabolic strategy.

4.2 Chlorenchyma Arrangement and Internal CO₂ Pathways

During Phase III CO₂ levels inside the leaf can get to about three up to ten times what's in the atmosphere, because malate decarboxylates in the cytoplasm. CAM plants handle this kinda high-CO₂ interior in part by how their photosynthetic tissue is set up. Chlorenchyma cells, the green photosynthetically active ones, are usually packed quite densely without much empty space between them. That helps lower the chances of CO₂ escaping back out through the stomata during Phase III, so internal carbon refixation becomes more effective.

Also, the wax like cuticle that covers most succulent leaves contributes. It's exceptionally impermeable, so cuticular transpiration (meaning water loss straight through the leaf surface rather than via stomata) stays at levels that are basically negligible compared with stomatal transpiration. Nobel (2003) pointed out that cuticular conductance in cacti and agaves is among the lowest recorded for any plant group, which basically supports the idea that these species can keep water balance steady even when desert conditions turn extreme.

4.3 The Role of PEP Carboxylase Regulation

One thing that makes CAM biochemistry genuinely fascinating is how tightly the PEPC enzyme is regulated, to stop futile cycling from happening. During the day, when CO₂ is being released from malate inside the cell, you don't really want PEPC running again and re-fixing that CO₂ into more malate... it would waste energy, and kind of defeat the whole point of releasing it toward RuBisCO. The plant manages this by the phosphorylation state of PEPC. At night, there's a dedicated kinase that phosphorylates PEPC, turning it on so CO₂ fixation can proceed. During the day, a phosphatase dephosphorylates it, so PEPC activity drops, and its sensitivity to malate inhibition also reduces.

And that circadian controlled enzyme adjustment is basically coordinated by the plant's internal clock. Research by Hartwell et al. (1999), published in *The Plant Journal*, identified the kinase responsible for nighttime PEPC activation and also confirmed that the regulation depends on the circadian system, so CAM isn't just some passive reaction to darkness, but an actively clocked biochemical program.

V. Ecological Diversity of CAM Plants

5.1 Which Families Employ CAM?

CAM shows up all over the plant tree of life, kind of scattered but with real striking breadth. Silvera et al. (2010) estimated CAM happens in around 7% of vascular plant species, across more than 60 families. The catalog includes a bunch of the most recognizable desert plants — cacti (Cactaceae), agaves (Asparagaceae), aloes (Asphodelaceae), stonecrops (Crassulaceae), and ice plants (Aizoaceae). Still, it also covers epiphytic orchids, bromeliads that live on rainforest tree branches, and even aquatic plants that deal with a different type of CO₂ restriction.

The unifying thread isn't really "desert," it's CO₂ limitation plus water stress. Epiphytic orchids in humid forests get limited CO₂ because their roots don't connect well to soil, so they become CO₂-limited compared with their light. Aquatic plants can also run into CO₂ limitation, since dissolved CO₂ just diffuses slowly through water. CAM basically fixes a concentration and timing problem, and honestly that problem shows up in more environments than most people think.

5.2 Facultative CAM and Metabolic Flexibility

Not all CAM plants are fully committed to the strategy full time. Some species end up switching between C₃ and CAM metabolism, depending on how much water is around, this kind of happening is usually called facultative CAM. When water is plentiful, they photosynthesize in a more regular way, the stomata open during the day and CO₂ gets fixed directly. Then when drought stress shows up they switch over into CAM mode. *Mesembryanthemum crystallinum*, the common ice plant, is like the most studied example where this shift happens.

As in Figure 2, the move from C₃ to CAM in *M. crystallinum* comes with a clear increase in PEPC activity and also a buildup of malate at night, within days after drought or salinity stress is imposed. Cushman and Bohnert (1999) described the molecular underpinnings behind this transition, pretty thoroughly, and they showed that stress responsive gene expression causes the upregulation of CAM specific enzymes, and in effect reprograms what the leaf is doing metabolically, as if its identity changes.

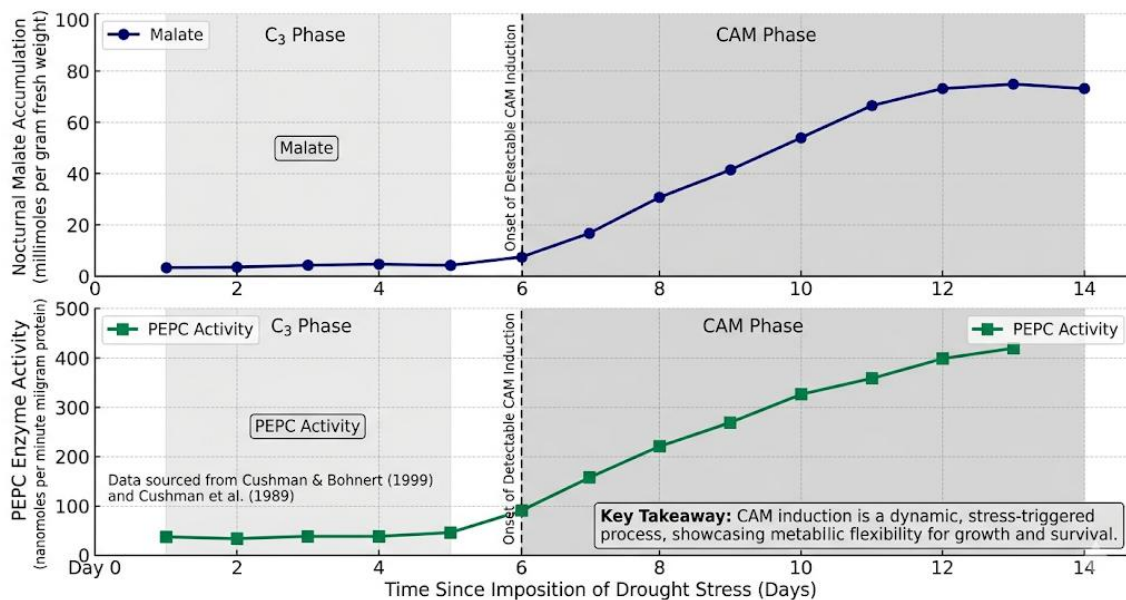


Figure 2: Induction of CAM Metabolism in *Mesembryanthemum crystallinum* Under Increasing Water Stress

This figure shows a line graph with two panels, one above the other. The upper panel tracks nocturnal malate accumulation, in millimoles per gram fresh weight, and the lower panel follows PEPC enzyme activity, in nanomoles per minute per milligram protein. The measurements come from *Mesembryanthemum crystallinum* leaves taken over 14 days after drought stress is imposed, meaning irrigation is withheld starting on day 0.

What you can see is that both variables stay low and kind of steady through days 1 to 5, and then they begin to climb in a more noticeable way starting from day 6. By days 12 to 14 the values are in the range you'd expect for constitutive CAM plants. There's also a vertical dashed line placed at day 6, this marks when CAM induction becomes detectable. The data is sourced from Cushman & Bohnert 1999 in *Annual Review of Plant Physiology and Plant Molecular Biology*, and from Cushman et al. 1989 in *Plant Cell*.

The main point is that CAM induction in facultative species looks like a shifting, stress driven event not like some fixed genetic thing. It illustrates how metabolic flexibility within the same plant can support both growth efforts and survival modes, depending on what the environment demands.

5.3 Water Use Efficiency: The Quantitative Advantage

The water saving advantages of CAM are not small, or marginal in any meaningful way. Nobel (1988) put together compiled data, and it suggests that CAM plants usually fix somewhere between 1 and 3 millimoles of CO₂ per mole of water transpired, while C₃ plants fall in a similar range of 1–3 millimoles of CO₂ per mole of water. But the key part, the crucial difference is how that number is obtained, for CAM it is done during the cooler night time, with a VPD that is less stressful and not as demanding. Then when people report water use efficiency at the whole plant, whole season level, CAM plants often land at values that are three to six times higher than C₃ plants that are growing in the same environment, Nobel (1991).

This point isn't just "basic ecology" talk either. Agricultural water use is about 70% of global freshwater withdrawals, according to the Food and Agriculture Organization of the United Nations (FAO, 2020). Crops that can keep productivity going even with lowered water inputs have huge practical value. Agave species, which are already grown for fiber and drink production across arid zones of Mexico and the American Southwest, have started getting more attention as drought tolerant bioenergy feedstocks, specifically because of their CAM related efficiency. Yang et al. (2015) modeled the global potential for agave productivity, and they concluded that dry and semi dry lands that don't work well for conventional crops could still produce notable agave yields, and their water needs are far less than those of corn or sugarcane.

VI. Conclusion

The question this article started with— how do desert succulents avoid starving for CO₂ without opening their stomata during the day — turns out to have a beautifully intricate answer, and also a kind of elegant messiness to it. These plants don't really "avoid" the problem so much as they restructure their whole biochemical timing around it. They gather their CO₂ at night, store it as malic acid inside their vacuoles, and then use it during the day to power photosynthesis. The result is a plant that can keep its stomata closed through

the harshest stretch of desert hours, while still making the sugars it needs to grow, reproduce, and just keep going.

What's striking about CAM is that it isn't some compromise or a half-measure. It's a full metabolic reorganization, one that includes coordinated shifts in enzyme regulation, cell layout, vacuolar capacity, cuticle properties, and circadian timing. Every part seems to support the others, rather than acting alone. The big vacuoles allow more malate storage. The less permeable cuticle makes stomatal control the main switch for water loss, essentially. Clock-driven enzyme activity also helps stop wasteful futile cycling. And the tightly packed chlorenchyma reduces internal CO₂ leakage during Phase III.

This level of integration is why CAM evolved sort of independently in so many lineages. The evolutionary pressure was real, and kind of severe. And the solution space, even if it stays biochemically complex, somehow converged on similar designs across wildly different plant families. There's something worth appreciating in that, the same logical answer to the same physical problem kept showing up again and again across hundreds of millions of years of plant evolution, like almost stubbornly consistent.

For researchers thinking about drought tolerant crops or water efficient food systems, CAM gives a proven, existing blueprint. The hard part is translating that blueprint into species and contexts where it doesn't naturally occur. Whether it becomes practically achievable is still an open question, but getting what succulents do to solve the CO₂ problem, without basically dying of thirst, is the essential first step.

References

- [1]. Borland, A. M., Griffiths, H., Hartwell, J., & Smith, J. A. C. (2009). Exploiting the potential of plants with crassulacean acid metabolism for bioenergy production on marginal lands. *Journal of Experimental Botany*, 60(10), 2879–2896. <https://doi.org/10.1093/jxb/erp118>
- [2]. Borland, A. M., Hartwell, J., Weston, D. J., Schlauch, K. A., Ferrier, J. M., Borland, J., Cushman, J. C., & Hartwell, J. (2014). Engineering crassulacean acid metabolism to improve water-use efficiency. *Trends in Plant Science*, 19(5), 327–338. <https://doi.org/10.1016/j.tplants.2014.01.006>
- [3]. Cushman, J. C., & Bohnert, H. J. (1999). Crassulacean acid metabolism: Molecular genetics. *Annual Review of Plant Physiology and Plant Molecular Biology*, 50, 305–332. <https://doi.org/10.1146/annurev.arplant.50.1.305>
- [4]. Cushman, J. C., Meyer, G., Michalowski, C. B., Schmitt, J. M., & Bohnert, H. J. (1989). Salt stress leads to differential expression of two isogenes of phosphoenolpyruvate carboxylase during crassulacean acid metabolism induction in the common ice plant. *Plant Cell*, 1(7), 715–725. <https://doi.org/10.1105/tpc.1.7.715>
- [5]. Dodd, A. N., Salathia, N., Hall, A., Kévei, E., Tóth, R., Nagy, F., Hibberd, J. M., Millar, A. J., & Webb, A. A. R. (2005). Plant circadian clocks increase photosynthesis, growth, survival, and competitive advantage. *Science*, 309(5734), 630–633. <https://doi.org/10.1126/science.1115581>
- [6]. Food and Agriculture Organization of the United Nations. (2020). *The state of the world's land and water resources for food and agriculture: Systems at breaking point*. FAO. <https://doi.org/10.4060/cb7654en>
- [7]. Hartwell, J., Smith, L. H., Wilkins, M. B., Jenkins, G. I., & Nimmo, H. G. (1999). Higher plant phosphoenolpyruvate carboxylase kinase is regulated at the level of translatable mRNA in response to light or a circadian rhythm. *The Plant Journal*, 20(3), 333–342. <https://doi.org/10.1046/j.1365-313x.1999.00609.x>
- [8]. Intergovernmental Panel on Climate Change. (2021). *Climate change 2021: The physical science basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press. <https://doi.org/10.1017/9781009157896>
- [9]. Lüttge, U. (2004). Ecophysiology of crassulacean acid metabolism (CAM). *Annals of Botany*, 93(6), 629–652. <https://doi.org/10.1093/aob/mch087>
- [10]. Nimmo, H. G. (2000). The regulation of phosphoenolpyruvate carboxylase in CAM plants. *Trends in Plant Science*, 5(2), 75–80. [https://doi.org/10.1016/S1360-1385\(99\)01543-5](https://doi.org/10.1016/S1360-1385(99)01543-5)
- [11]. Nobel, P. S. (1988). *Environmental biology of agaves and cacti*. Cambridge University Press.
- [12]. Nobel, P. S. (1991). Achievable productivities of certain CAM plants: Basis for high values compared with C₃ and C₄ plants. *New Phytologist*, 119(2), 183–205. <https://doi.org/10.1111/j.1469-8137.1991.tb01022.x>
- [13]. Nobel, P. S. (2003). *Cacti: Biology and uses*. University of California Press.
- [14]. Osmond, C. B. (1978). Crassulacean acid metabolism: A curiosity in context. *Annual Review of Plant Physiology*, 29, 379–414. <https://doi.org/10.1146/annurev.pp.29.060178.002115>
- [15]. Pittermann, J., & Sperry, J. S. (2003). Tracheid diameter is the key trait determining the extent of freezing damage in conifers. *Tree Physiology*, 23(13), 907–914. <https://doi.org/10.1093/treephys/23.13.907>
- [16]. Silvera, K., Neubig, K. M., Whitten, W. M., Williams, N. H., Winter, K., & Cushman, J. C. (2010). Evolution along the crassulacean acid metabolism continuum. *Functional Plant Biology*, 37(11), 995–1010. <https://doi.org/10.1071/FP10084>
- [17]. Winter, K., & Smith, J. A. C. (Eds.). (1996). *Crassulacean acid metabolism: Biochemistry, ecophysiology and evolution*. Springer. <https://doi.org/10.1007/978-3-642-79060-7>
- [18]. Yang, X., Cushman, J. C., Borland, A. M., Edwards, E. J., Wulfschleger, S. D., Tuskan, G. A., Owen, N. A., Griffiths, H., Smith, J. A. C., De Paoli, H. C., Weston, D. J., Cottingham, R., Hartwell, J., Davis, S. C., Silvera, K., Ming, R., Schlauch, K., Abraham, P., Stewart, J. R., ... Cushman, J. C. (2015). A roadmap for research on crassulacean acid metabolism (CAM) to enhance sustainable food and bioenergy production in a hotter, drier world. *New Phytologist*, 207(3), 491–504. <https://doi.org/10.1111/nph.13393>
- [19]. Zhu, J.-K. (2016). Abiotic stress signaling and responses in plants. *Cell*, 167(2), 313–324. <https://doi.org/10.1016/j.cell.2016.08.029>